

LABINC
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Bioinorgânica e Cristalografia

“Bioinorganic Catalysis”

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Research in our laboratories is concerned with the design of symmetrical and **unsymmetrical dinuclear small molecules** as structural and functional models for the active-site of metalloenzymes including **hydrolases (PAPs), proteases, and catechol oxidases**.

rkbPAP
Klabunde et al.
J. Mol. Biol. 259 (1996) 737

Sp Catox
Krebs et al. *Nature*,
5 (1998) 1084

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PAPs – catalyze - **hydrolysis** - phosphorylated substrates - pH range 4-7.

Biological roles of PAPs:
Mammalian - Iron transport during pregnancy and macrophage-specific generation of ROS. Associated with metabolic bone diseases (osteoporosis, cancers with metastases).
Plant - May play a crucial role in immobilizing organic phosphate esters in the soil during germination.

PAP enzymes are targets for the design of drugs for a wide variety of disorders

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Catechol Oxidase

- ✓ **Catechol oxidase** - plant enzyme - belongs to the oxidoreductase class.
- ✓ **Active site** - Dinuclear Copper center .
- ✓ **Catalyzes oxidation of o-diphenols** (like caffeic acid) to the corresponding o-quinones, with the reduction of molecular oxygen to water.
- ✓ The highly reactive quinones auto-polymerize to form brown polyphenolic catechol melanins, a process thought to protect the damaged plant from pathogens or insects.
- Reaction of great importance** in the medical diagnosis of hormonal catechol amines (adrenalin, noradrenalin, and dopamine).

In spite of extensive studies on the mechanism of CatOx the coordination mode of the substrate remains unknown.

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Catalytic Promiscuity

Catalytic promiscuity means, in the broadest terms, the ability of a given active site to catalyze more than one chemical transformation and constitutes a very important property of many enzymes, having a natural role in evolution and, occasionally, in the biosynthesis of secondary metabolites. Important role in the diversification of enzymes by providing a duplicated gene a head start towards being captured by adaptive evolution.

R. J. Kazlauskas, *Curr. Opin. Chem. Biol.* 9 (2005)195-201.

Ex.

Acid phosphatase from plant uniquely exhibits chloroperoxidase activity with loss of phosphatase activity when orthovanadate is added to the apo form of the enzyme.

Carbonic anhydrase also catalyses the hydrolysis of triesters – **phosphotriesterase activity**.

Important: Very well known phenomena for a diversity of enzymes. However, very few examples are described for models.

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Strategy Utilized – Synthetic Hydrolase

(metal ion at the catalytically active site)

- Activate the substrate and/or stabilize the transition state
- Liberate products at a reasonable rate
- Reduce pK_a of coordinated H₂O (provide nucleophiles at ≈ neutral pH)

Use of unsymmetrical ligands – mimicking most natural metalloenzymes

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Synthesis of the unsymmetrical hetero-dinuclear FeZn and FeCu complexes:

H₂L-R and H₂py₂mpf

1 - Zn(ClO₄)₂·6H₂O
2 - Fe(ClO₄)₃·9H₂O
3 - NaOH/MeOH

**[L(OH)₂Fe^{III}(μ-OH)Zn^{II}](ClO₄)₂
FeZnL-R**

1 - Cu(ClO₄)₂·6H₂O
2 - Fe(ClO₄)₃·9H₂O
3 - NaOH/MeOH

**[L(OH)₂Fe^{III}(μ-OH)Cu^I](ClO₄)₂
FeCuL-R**

Schenk, Neves et al., *Chem. Rev.* 2006, 106, 3338-3363 Fe^{III}(OAc)₂M^{II} (M^{II} = Fe, Mn, Zn, Cu, Ni, Co)
Inorg. Chem. 2002, 41, 4624; Ga^{III}(OAc)₂Zn^{II}
Inorg. Chem. 2002, 41, 5641;
Inorg. Chem. Commun. 2002, 5, 434 and 2003, 6, 1161;
J. Biol. Inorg. Chem. 2005, 10, 319; *J. Biol. Inorg. Chem.* 2008, 13, 139; *J. Biol. Inorg. Chem.* 2007, 12, 1207

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H₂L-R R = H, NO₂, CH₃

Zn-Fe 3.040(5)
Zn-O(1)-Fe 96.2(3)
Zn-O(1)-Fe 102.4(3)

Cu-Fe 3.020(5)
Cu-O(1)-Fe 96.2(3)
Cu-O(2)-Fe 102.9(3)

The structure is stabilized by H-contacts:
1) Bridging OH with oxygen of ClO₄⁻
2) Terminal Fe-OH₂ with O1W;

$\tau = (\alpha - \beta)/60 = 0.71$ for Zn^{II} 0.70 for Cu^I
- Addition -
- distorted trigonal bipyramidal

Neves et al., *JACS*, 2007, 129, 7486
First [Fe^{III}(μ-OH)Zn^{II}]

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Comparison of the structures of FeZnL-H and kbPAP

kbPAP

FeZnL-H

Important: Both structures contain an Fe^{III}(μ-OH)Zn^{II} with Fe...Zn distances of 3.05 Å in the FeZnL-H complex and 3.2 Å in kbPAP – Krebs, Witzel et al., *J. Mol. Biol.* 1996, 259, 737.

Both structures also contain terminal Fe^{III}-phenolate and Fe^{III}-OH bonds

Neves et al., *JACS*, 2007, 129, 7486

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Hydrolytic Activity in the Hydrolysis of BDNPP

✓ pH dependence

Active species

FeZnL-H – optimum at 6.5
pK₁ = 5.0 and pK₂ = 8.1

FeCuL-H – optimum at 7.0
pK₁ = 5.3 and pK₂ = 8.6

Active species under physiological conditions

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Dependence of the catalytic activity on [2,4-BDNPP] – pH optimum

FeZnL-H

FeCuL-H

186 turnovers in 24 h;
[FeZnL-H] = 5x10⁻⁷M

Complex	K _{ass} (L.mol ⁻¹)	K _M (mol.L ⁻¹)	k _{cat} (s ⁻¹)	k _{cat} /K _M
FeZnL-H pK = 4.66	238	4.20 x 10 ⁻³	9.13 x 10 ⁻⁴	4830
FeCuL-H pK = 5.23	117	8.52 x 10 ⁻³	19.0 x 10 ⁻⁴	10100

$V_0 = V_{max} [S]/(K_m + [S])$
 $1/V_0 = K_m/V_{max}[S] + 1/V_{max}$

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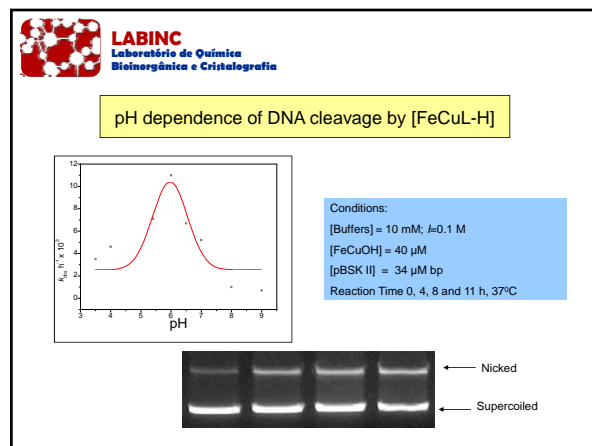
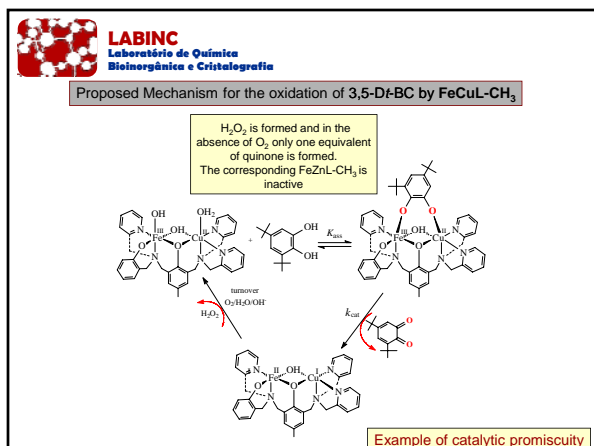
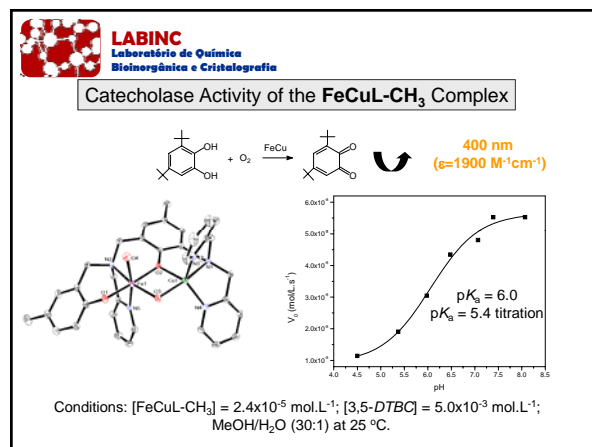
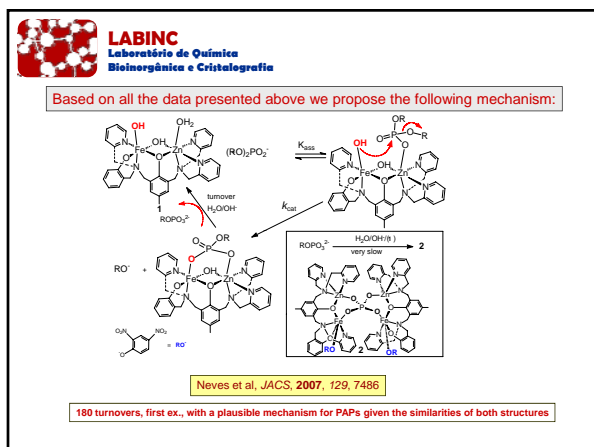
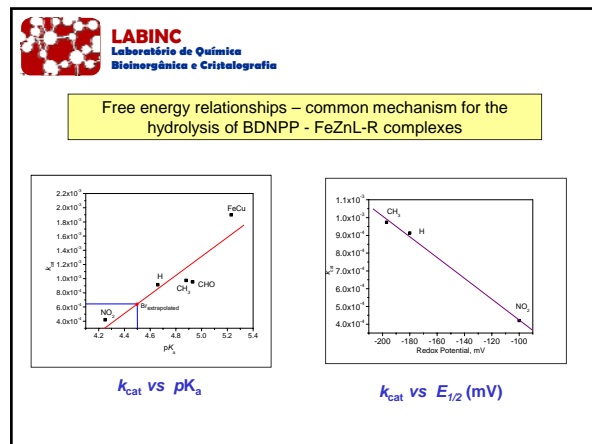
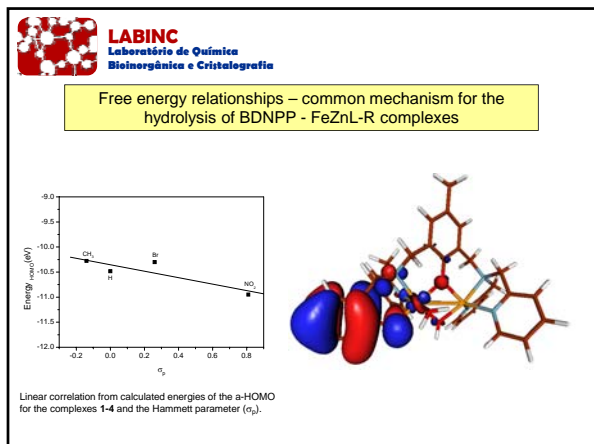
Kinetics parameters for FeZnL-R and FeCuL-H

Complex	pK ₁	K _{ass} (L.mol ⁻¹)	E = k _{cat} /K _M (mol ⁻¹ .L.s ⁻¹)	k _{cat} (s ⁻¹)	E _{1/2} mV vs NHE (pH 6.0)	k _{cat} /K _M
FeZnL-CH ₃	4.88	192	19 x 10 ⁻²	9.74 x 10 ⁻⁴	-197	5.2 x 10 ³
FeZnL-H	4.66	238	20 x 10 ⁻²	9.13 x 10 ⁻⁴	-180	4.8 x 10 ³
FeZnL-Br	4.50	168	11 x 10 ⁻²	6.55 x 10 ⁻⁴	-160	3.5 x 10 ³
FeZnL-NO ₂	4.25	286	12 x 10 ⁻²	4.20 x 10 ⁻⁴	-100	2.2 x 10 ³
FeCuL-H	5.23	117	85 x 10 ⁻²	19.0 x 10 ⁻⁴	-430	10.0 x 10 ³
FeZnpy ₂ mpf	4.93	67	6 x 10 ⁻²	9.53 x 10 ⁻⁴	-252	5.1 x 10 ³

Kinetic isotope effect k_H/k_D = 1.00 – 1.34

EXAFS studies - in CH₃CN/H₂O solution reveal a Fe^{III}-Zn^{II} bond distance of 3.043 Å for FeZnL-H in full agreement with the X-ray structure

R = H, CH₃, NO₂



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Dependence of the DNA Cleavage on [FeCuL-H]

Under pseudo-Michaelis-Menten conditions the following parameters were obtained: $k_{cat} = 0.98 \text{ h}^{-1}$; $t_{1/2} = 42 \text{ min}$; $K_m = 3.9 \times 10^{-5} \text{ M}$, Providing a rate enhancement of 2.7×10^7 over the uncatalyzed reaction thus being one of the most active described to date.

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Cytotoxicity Studies on cancer Cells

Fe(μ-OH)Zn^{II}

Complex	IC ₅₀ (μM)
1 - CH ₃	5.73±06
2 - H	6.2±05
3 - Br	21.9±2.0
4 - NO ₂	47.9±4.0

It is worth noting that cytotoxic activity increases in the same order as the hydrolysis of 2,4-bdnpp and DNA.

A. Neves, et al. *J. Biol. Inorg. Chem.* 2009 submitted.

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Innovation - New Projects Underway in the Group

Development of new bioinorganic-materials with chitosan as drug-delivery systems

Non-toxic natural material

Complex already synthesized and fully characterized

Heterogeneous Catalysis

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Heterogeneous Catalysis

Piovezan and Neves, Manuscript in Preparation

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Complex	V ₀ (mol.L.s ⁻¹)	K _M (mol/L)	k _{cat} (s ⁻¹)	*K ₁₅ (L/mol)	*E (L.s/mol)	F (K _{cat} /K _{uncat})	Turn over (1h)
NINlppamif	5.37x10 ⁻⁷	1.57x10 ⁻³	5.4x10 ⁻²	637	34.2	298.300	33.4
Si3AP-NINI	9.20x10 ⁻⁷	7.30x10 ⁻⁴	3.5x10 ⁻²	1370	48.2	195.500	78.5
Si3APTS-NINI	3.40x10 ⁻⁷	6.9x10 ⁻⁴	4.6x10 ⁻²	1450	66	255.500	32.7
Simag-NINI	4.19x10 ⁻⁷	1.21x10 ⁻³	3.5x10 ⁻²	826	29	196.650	23

*K₁₅ = 1/K₁₀; *E = K₁₀K₁₅ (catalytic efficiency).
at pH 9.0
Si3AP – silica 3-aminopropyl
Si3APTS – silica 3-aminopropyl triethoxisilano
Simag – magnetic particle recovered with silica

Piovezan and Neves, Manuscript in Preparation

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Suitable for Catalysis as chemical Nuclease or catecholase

The second coordination sphere becomes much more important

Intercalates with DNA and shows Fluorescent properties

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Very recent results

Preliminar results reveal that the Cu^{II}Cu^I complex of L1a catalyses the oxidation of 3,5-di-tert-butylcatechol with high efficiency and formation of H₂O as observed in the enzyme catechol oxidase. The complex also catalyses the hydrolysis of DNA and proteins (BSA)
Catalytic promiscuity and possibility to solve the mechanism of catechol oxidase

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**Facilities available in our Labs
in Collaboration with other Groups**

- 1) **X-ray structure determinations.** (UFSC - Florianópolis/Brazil – Profs. Ademir Neves & Adailton Bortoluzzi - New CCD available up from 2010).
- 2) **EXAFS – structure in solution.** (Australia – Prof. Dr. G. Schenk).
- 3) **Magnetic susceptibility and MCD** in the solid state and in solution between 4.2 e 350 K (Germany– Prof. Dr. W. Haase).
- 4) **HFEPR** measurements. (USA - Prof. Dr. Joshua Telser).
- 5) **DNA and protein interactions.** (UFSC - Florianópolis/Brazil – Profs. Hernan Terenzi & Ademir Neves).
- 6) **Cytotoxic Activity.** (UFMG – Prof. Dr. Elene Pereira Maia).

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O grupo unido, jamais será vencido

